

Manual non-radioactive RNA *in situ* hybridization steps with amplification strategy

Tissue: fresh frozen E14.5 embryo and adult brain sections (PFA-fixed, acetylated and dehydrated)

Day 1

<i>Incubate</i>		Methanol + 3% H ₂ O ₂	20 min	RT
<i>Wash</i>		PBS	5 min	RT
<i>Denature</i>	0.2 N	HCl	10 min	RT
<i>Wash</i>		PBS	2 x 5 min	RT
<i>Deproteinase</i>		PK-buffer + 5-10 µg/ml proteinase K ¹	2 x 10 min	RT
<i>Wash</i>		PBS	2 x 5 min	RT
<i>Fix</i>	4 %	PFA in PBS	15 min	RT
<i>Wash</i>		PBS	2 x 5 min	RT
<i>Pre-hybridize</i>		HyB-Buffer (add 1mg/ml DTT)	30 min	60 °C
<i>Hybridize</i>		HyB-Buffer + 0.5-2µg/ml probe (parafilm)	o/n	60 °C

Day 2

<i>Wash</i>	2x	SSC	15 min	60 °C
<i>Wash</i>	2x	SSC	5 min	RT
<i>Wash</i>	0.2x	SSC	2 x 30 min	60 °C
<i>Wash</i>	0.2x	SSC	2 min	RT
<i>Wash</i>		TNT buffer (0.1% Tween 20 in TN)	2 x 20 min	RT
<i>Block</i>	10 %	inactivated sheep serum (SS) in TNB	30 min	RT
<i>Antibody</i>	10 %	SS in TNB + anti-DIG (POD) antibody (1:500)	30 min	RT
<i>Wash</i>		TNT buffer (0.1% Tween 20 in TN)	2 x 20 min	RT
<i>Incubate</i>		Tyramide Biotin (1:50 conc.)	20 min	RT
<i>Wash</i>		Maleate wash buffer	2 x 20 min	RT
<i>Incubate</i>		Neutravidin-AP	30 min	RT
<i>Wash</i>		Maleate wash buffer	2 x 20 min	RT
<i>Wash</i>		TMN (pH 9.5; add 5mM Levamisole freshly)	2 x 5 min	RT
<i>Stain</i>		TMN + 1µl/ml NBT and 3.5µl/ml BCIP	3 x 15 min ²	RT dark

Day 3

<i>Wash</i>	PBS	2 x 5 min	RT
<i>Fix</i>	4 % PFA	2 x 5 min	RT
<i>Wash</i>	PBS	2 x 5 min	RT
<i>Mount</i>	Hydro-Mount (cover slipped)	> 120 min	37 °C

¹The required proteinase K concentration depends largely on the tissue type. By default we use 5µl/100ml of proteinase K (Roche 3115828) for E14.5 mouse embryo and 35µl/100ml for adult mouse / monkey brain.

²**Critical:** Check the signal intensity every 10 mins during the last staining step. Higher expressed genes will be stained within 10 mins, whereas lower expressed genes might take 30 – 35 mins.