Primer Design

- 1. Go to NCBI (National Center for Biotechnology Information) website (<u>National Center for</u> <u>Biotechnology Information (nih.gov)</u>
- 2. On the upper part of the page you will find:



3. Click on All Databases



4. Select 'Gene'



5. Type the name of your gene of interest in the query box (GFAP for example)



6. Find your species of interest. Click ctrl+F and type in the name of the species (in our case this is *Macaca mulatta (Rhesus monkey)*

| GFAP ID: 419969 | glial fibrillary acidic protein [<i>Gallus gallus</i> (chicken)] | Chromosome 27, NC_006114.5 (36976283702163, complement) | |
|-----------------------|--|--|----------------|
| GFAP ID: 396562 | glial fibrillary acidic protein [<i>Sus scrofa</i> (pig)] | Chromosome 12, NC_010454.4 (1845626818466594) | |
| GFAP ID: 712941 | glial fibrillary acidic protein [Macaca mulatta (Rhesus monkey)] | Chromosome 16, NC_041769.1 (5616938956180984, complement) | |
| GFAP ID: 454741 | glial fibrillary acidic protein [<i>Pan troglodytes</i> (chimpanzee)] | Chromosome 17, NC_036896.1 (1032305710333051) | CK820_G0052909 |
| GFAP ID: 100033970 | glial fibrillary acidic protein [<i>Equus caballus</i> (horse)] | Chromosome 11, NC_009154.3 (1881510118824260) | |
| GFAP ID: 101081938 | glial fibrillary acidic protein [<i>Felis catus</i> (domestic cat)] | Chromosome E1, NC_018736.3 (4486846844877976, complement) | |
| GFAP ID: 101017460 | glial fibrillary acidic protein [<i>Papio anubis</i> (olive baboon)] | Chromosome 17, NC_044992.1 (5172307951734689, complement) | |
| GFAP ID: 101339778 | glial fibrillary acidic protein [<i>Tursiops</i> <i>truncatus</i> (common bottlenose dolphin)] | Chromosome 20, NC_047053.1 (4100782441018040, complement) | |
| Gfap ID: 100758767 | glial fibrillary acidic protein [Cricetulus | | 179_011607 |
| | | | |

7. Click on the gene name. In the following page you should find *mRNA and Protein(s)* section

| enomic | | | | |
|--------------------------|---|--|-----|--|
| | | | | |
| 1. NC_041769.1 Reference | e Mmul_10 Primary Asse | nbly | | |
| Range | 5616938956180984 compl | ement | | |
| Download | GenBank, FASTA, Sequence | Viewer (Graphics) | | |
| RNA and Protein(s) | | | | |
| | | | | |
| 1. XM_015119892.2 → XF | 014975378.1 glial fibrilla | ry acidic protein isoform X2 | | |
| Related | ENSMMUP00000049963.2, | ENSMMUT00000076046.2 | | |
| Conserved Domains (2) st | mmary | | | |
| | pfam00038 Location:138 → 446 | Filament; Intermediate filament protein | | |
| | $\frac{pfam04732}{Location:73 \rightarrow 136}$ | Filament_head; Intermediate filament head (DNA binding) region | | |
| 2. XM_028836604.1 → XF | 028692437.1 glial fibrill | rry acidic protein isoform X1 | | |
| Related | ENSMMUP00000046183.2, | ENSMMUT00000077212.2 | | |
| Conserved Domains (2) st | mmary | | | |
| | <u>pfam04732</u> Location:71 → 136 | Filament_head; Intermediate filament head (DNA binding) region | | |
| | pfam00038 Location:138 → 446 | Filament; Intermediate filament protein | | |
| | | | | |
| ted sequences | | | ≈ ? | |
| | | | | |

- Click <u>XM_015119892.2</u> which is accession number of the mRNA sequence N.B. <u>XP_014975378.1</u> is the protein sequence ; prefix XM_ (mRNA), XR_ (non-coding RNA), and XP_ (protein)
- 9. The following page will show you the all the information about the mRNA of the gene that you are searching.
- 10. Next click FASTA to obtain the full sequence of the mRNA of your gene of interest.

| S NCBI F | Resources 🗹 How To 🕑 | | Sign in to NCE |
|--------------------------------|---|------------|--|
| Nucleotide | e Nucleotide Advanced | | Search |
| GenBank - | | Send to: - | Change region shown |
| PREDIC variant | CTED: Macaca mulatta glial fibrillary acidic protein (GFAP), transcript X2, mRNA | | Customize view |
| NCBI Refere | ance Sequence: XM_015119892.2 | | Analyze this sequence Run BLAST |
| <u>🕫 ta</u> 🖂 | | | Pick Primers |
| LOCUS | XM_015119892 4016 bp mRNA linear PRI 26-APR-2019 | | Highlight Sequence Features |
| DEFINITION | transcript variant X2, mRNA. | | Find in this Sequence |
| ACCESSION VERSION DBLINK | XM_015119892 XM_015119892.2 BioProject: PRJNA528504 Padera | | Show in Genome Data Viewer |
| SOURCE | Narosay Macaca mulatta (Rhesus monkey) <u>Macaca mulatta</u> Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglira; Perimates; Haplorrhini; Granchini; Garconitheridae; Garconitherinae; Macaca | | Articles about the GFAP gene A new rhesus macaque assembly and annotati for next-generation sequencing [Biol Direct. 201 |
| COMMENT | Calain many, demonstrates, cercoprinting ratata: MODEL REFSEQ: This record is perioticed by automated computational analysis. This record is derived from a genomic sequence (KC_041750, 1) annotated using gene prediction method: Gnomon, supported by mRNA and EST evidence. | | Age-related decreases in SYN levels associate with increases in MAP-2, apo [Age (Dordr). 20' See a |
| https://www.pcbi | Also See: Documentation of NCBI's Annotation Process | | Pathways for the GFAP gene |

- 11. Copy the sequence and transfer it into a word file
- 12. Go to Primer3 Input website
- **13.** Paste the sequence in the box

| Primer3web version 4.1.0 - | disclaimer cautions | <u>code</u> | | | | | | |
|---|---|---------------------|--|------------------------|------------|--|--|--|
| Select the <u>Task</u> for primer selection generic | v | | | | | | | |
| Template masking before primer design (ava | ilable species) | | | | | | | |
| Select species Example: Mus musculus | Nucleotides to mask in 5' direction 1 | | | | | | | |
| Primer failure rate cutoff < 0.1 | Nucleotides to mask in 3' direction 0 | | | | | | | |
| Paste source sequence below (5'->3', string o ALUs, LINEs, etc.) or use a <u>Mispriming Libr</u> | f ACGTNacgtn other letters treated as N - rary (repeat library) NONE | - numbers and blank | s ignored). FASTA format ok. Please N-ou | t undesirable sequence | e (vector, | | | |
| GGTAGTGCCATATTTTACAATTTGTAAAACAAGCACGAG GCCTGGAGGCCGGGTGCTCAGGGCTGACATGTCCACCCC | CAAAATGAAGACACTGGCTCATATTCCTGCA AGTGCACCCACTCTGCTTTTTAACTGGGCAG | | • | | | | | |
| ACTOGTAAGCAGACTGGTGGGATCTGTGCCCAGAAATGG TCCCCTCTAAGGCTGAAGAAGGGTCCTTCCCCCTCCCCA | AGACTGGGAGGGCCCACTTCAGGGTTCTCCTC | | | | | | | |
| TGCCACCTGCTGCTGCTGCTGCTGCTGCTATCTTCAGGGCA AAAGACACATGCTGCGCCCTTCCCCA | CTGCTGCTGCCTTTAGTCGCTGAGGAAAAAT | | | | | | | |
| Pick left primer. | Pick hybridization probe (internal | | Pick right primer, or use right primer h | elow |] | | | |
| or use left primer below | oligo), or use oligo below | | (5' to 3' on opposite strand) | | | | | |
| | | | | | | | | |

14. Next go down and find General Primer Picking Conditions. Using the information that we have provided in Primer Design Rules file, fill the spaces as indicated.

| Upload the settings from a | file | Choose File | No fi | ile chosen | | | | | |
|----------------------------|-------|-------------|--------|-----------------|-----------------------|--------------------|-----------------|-----------------|---|
| Primer Size Min 19 | Opt | 20 | Max | 21 |] | | | | |
| Primer Tm Min 58.0 | Opt | 59.0 | Max | 61 | Max Tm Difference 5.0 | Table of thermodyn | amic parameters | SantaLucia 1998 | v |
| Product Tm Min -10000 | 0 Opt | 0.0 | Max | 1000000. | | | | | |
| Primer GC% Min 45 | Opt | 50.0 | Max | 55 |] | | | | |
| | | | - | | | | | | |
| Product Size Ranges 700 | 300 | | | | | | | | |
| Number To Return | 5 | | | <u>Max 3' S</u> | tability 9.0 | | | | |
| Max Library Mispriming | 12.00 | Pair M | ax Lił | orary Mis | priming 20.00 | | | | |

N.B. Choose your product size range accordingly to the length of your mRNA sequence. Optimal range amplicon length product is between 600-1000bp. For genes with shorter than 600bp of length we usually design amplicon which ranges between 60-70% of the total length. Note that for those smaller gene there is no strict rule about the length of the amplicon.

15. Scroll down in the page, find, and click the button PICK PRIMERS.

General Primer Picking Conditions

11.12.2020

| \leftarrow \rightarrow \circlearrowright \textcircled{a} \textcircled{b} https: | ://prime | er3.ut.ee | | | -5 | 杀 | 5∕≡ | 回 | 6 | |
|--|---------------------|---|------------|---|-------|----|-----|---|---|--|
| M Gmail 🔹 YouTube 😝 Facebook 🔢 | •1 PDB-10 | D1 | | | | | | | | |
| Max GC in primer 3' end 5 | 5 | | · | | | | | | | |
| 3' End Distance Between Left Primers 3 | 3 | 3' End Distance Between Right Primers | 3 |] | | | | | | |
| 5 Prime Junction Overlap 7 | 7 | 3 Prime Junction Overlap | 4 | (Distance of the primer ends to one overlap pos | ition | .) | | | | |
| Concentration of Monovalent Cations 5 | 50.0 | Salt Correction Formula | SantaLuc | ia 1998 🗸 🗸 | | | | | | |
| Concentration of Divalent Cations | 1.5 | Concentration of dNTPs | 0.6 |] | | | | | | |
| Annealing Oligo Concentration 5 | 50.0 | (Not the concentration of oligos in the r | eaction mi | x but of those annealing to template.) | | | | | | |
| Sequencing Spacing 5 | 500 | Sequencing Interval | 250 |] | | | | | | |
| Sequencing Lead 5 | 50 | Sequencing Accuracy | 20 |] | | | | | | |
| Liberal Base Show Debug | ging Info | Treat ambiguity codes in libraries as | consensu | <u>s</u> | | | | | | |
| Lowercase masking Dick anyway | x | Print Statistics | | | | | | | | |
| Pick Primers Download Settings Rese Operative Function Penalty Weig | et Form ghts for | Primers | | | | | | | | |
| Size 1.0 Gt 1.0 Im Lt 1.0 Gt 1.0 GC% Lt 0.0 Gt 0.0 | | | | | | | | | | |
| TH: Self Complementarity 0.0 TH: 3' End Self Complementarity 0.0 TH: Hairpin 0.0 TH: Template Mispriming 0.0 | | | | | | | | | | |
| Self Complementarity 0.0 3' End Self Complementarity 0.0 | | | | | | | | | | |

16. The page that will open will show you the primes that the program picked

| Primer3 Output |
|--|
| PRIMER PICKING RESULTS FOR |
| Template masking not selected No mispriming library specified Using 1-based sequence positions |
| OLIGO start len tm gc% any_th 3'_th hairpin seq LEFT PRIMER 1775 20 59.14 55.00 3.32 0.00 0.00 GTTGGAGCTGACAGACG RIGHT PRIMER 2512 20 59.05 55.00 0.00 0.00 GTTGGAGTTTCTGGGTGGCTG SEQUENCE SIZE: 4016 INCLUDED REGION SIZE: 4016 |
| PRODUCT SIZE: 738, PAIR ANY_TH COMPL: 0.30, PAIR 3'_TH COMPL: 0.00 |
| 1 TGCTTTGTGGAGCTGTCAAGGCCTGGGCTCTGGGAAAGAGGCACAGGGAGGCCAGGCAAG |
| 61 GAAGGAGTGACCCGGAGGGACAGACCCAGGGGCTAAAGTCCTGATAGGGCAAGAGAGTGC |
| 121 CAGCCCCCTCTTGCTCTATAAGGACCTCCACTGCCACATACAGGCCATGATTGAT |
| 181 GACAAAGGGCTGGTGTCCAATCCCAGCCCCCAGAACTCCAGGGAATGAAT |
| 241 AGAGAGCAGGAATATGGGACATCTGTGTTCGAGGGGGGGG |
| 301 GGGCCTAGTAGGAAATGATGATATAGGCACCCCTTGAGGGTACTGAACAGGTTTGTTCTT |
| 361 CGCCAAATTCCCAGCACCTTGCAGGTACTTACAGCTGAGTGAAAGAAGGCCTGGGTTATG |
| 421 AAATCAAAAAGTTGGAAAGCAGGTCAGAGGTCATCTGGTACAGCCCTTCCTT |

You can see some information about the primers that the program picked.

- 1. In the red box (start) is the information about the position of the 5' base of the primer.
- 2. In the green box (len) is the information regarding the length of the primer.
- 3. In the yellow box (Tm) is the information regarding the melting temperature of the primer.
- 4. In the violet box (gc%) is the information about the GC% content of the primers or the percent of G or C bases in the primer.
- 5. In the orange box -any th (Self-Complementarity) and brown box -(hairpin) in the information about the tendency to primers to anneal to itself or to form secondary structure (hairpins). The lower the value the better.

Self-Complementarity 5' ACGTGCGTGAATACGT 3`



In the violet box (3`th)
 The self-complementarity of the primer at the 3` ends (primer dimers formation with itself). The lower the value the better.

- 7. The final Gray box (seq) represent the primers sequence.
- 8. Copy and paste all the information from the BLACK BOX into your word file.
- 9. Add T7 promotor sequence to the forward primer-

T7 GCGTAATACGACTCACTATAGGG + Forward PRIMER

10. Add Sp6 promotor sequence to the reverse primer-

Sp6 GCGATTTAGGTGACACTATAG + Reverse PRIMER