

## Manual non-radioactive RNA in situ hybridization

### Materials:

1. 1x PBS<sup>1</sup>
2. 0.2 HCL<sup>2</sup>
3. PK- buffer<sup>3</sup>
4. proteinase K (Roche 3115828)
5. 4% PFA<sup>4</sup>
6. Hyb-Buffer<sup>5</sup>
7. DTT (1,4-Dithiothreitol) (Merch 11583786001)
8. Parafilm
9. SSC<sup>6</sup>
10. PBST<sup>7</sup>
11. 10% Sheep-Serum- inactivated (Equitech-bio SSA62) <sup>8</sup>
12. TMN buffer<sup>9</sup>
13. Levamisole 5mM (Merck 1359302-125MG)
14. Anti-DIG (AP) antibody (Merck 11093274910)
15. Glass slides and coverslips and mounting media (water based)
16. RNA probe
17. NBT (Merck- 11383213001)
18. BCIP (Merck 11383221001)

### Method:

#### Manual non-radioactive RNA *in situ* hybridization

Tissue: fresh frozen tissue that are fixed in PFA, acetylated, and dehydrated <sup>10</sup>

#### Day 1

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<i>Wash</i>	PBS	5 min	RT
<i>Denature</i>	0.2 N HCl	10 min	RT
<i>Wash</i>	PBS	2 x 5 min	RT

<i>Deproteinase</i>		PK-buffer + 5-10 µg/ml proteinase K <sup>11</sup>	2 x 10 min	RT
<i>Wash</i>		PBS	2 x 5 min	RT
<i>Fix</i>	4 %	PFA in PBS	15 min	RT
<i>Wash</i>		PBS	2 x 5 min	RT
<i>Pre-hybridize</i>		HyB-Buffer (add 1mg/ml DTT)	30 min	60 °C
<i>Hybridize</i>		HyB-Buffer + 0.5-2µg/ml probe (parafilmmed)	o/n	60 °C

## Day 2

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<i>Wash</i>	2x	SSC	15 min	60 °C
<i>Wash</i>	2x	SSC	5 min	RT
<i>Wash</i>	0.2x	SSC	2 x 30 min	60 °C
<i>Wash</i>	0.2x	SSC	2 min	RT
<i>Wash</i>		PBST (0.1% Tween 20 in PBS)	2 x 20 min	RT
<i>Block</i>	10 %	inactivated sheep serum (SS) in PBST	30 min	RT
<i>Antibody</i>	10 %	SS in PBST + anti-DIG (AP) antibody (1:1000)	90 min	RT
<i>Wash</i>		PBST	2 x 20 min	RT
<i>Wash</i>		TMN (pH 9.5; add 5mM Levamisole freshly)	2 x 5 min	RT
<i>Stain</i>		TMN + 1µl/ml NBT and 3.5µl/ml BCIP	> 120 min <sup>12</sup>	RT dark

## Day 3

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<i>Wash</i>		PBS	2 x 5 min	RT
<i>Mount</i>		Hydro-Mount (cover slipped)	> 120 min	37 °C

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Notes:

1. 1x PBS solution preparation-  
[Phosphate-buffered saline \(PBS\) \(cshlp.org\)](#)
2. 0.2HCL solution preparation- To make a 0.2M solution, slowly add 16.42mL of your stock 37% solution of HCL to 250mL deionized H<sub>2</sub>O. Adjust the volume of the solution to 1000mL with Deionized water.  
N.B. Always add the hydrochloric acid to the water, not the water to the hydrochloric acid!  
For other calculation you can use this calculator-  
[Molarity Calculator & Normality Calculator for Acids & Bases | Sigma-Aldrich](#)
3. PK (proteinase K) buffer-  
[Proteinase K buffer \(10X\) \(cshlp.org\)](#). After you prepare 10X dilute the solution to 1X.
4. 4% PFA solution preparation-  
[Paraformaldehyde \(PFA; 4%\) \(cshlp.org\)](#)
5. Hyb-buffer alternative to the commercially ready-made solutions:  
[Hybridization buffer \(A\) \(cshlp.org\)](#) + [Denhardt's solution \(100X\) \(cshlp.org\)](#)
6. SSC solution preparation- [SSC \(cshlp.org\)](#).
7. PBST solution preparation- [PBST \(cshlp.org\)](#)
8. To inactivate the sheep serum heating to 56°C for 30 minutes, then store -20°C
9. TMN buffer solution preparation protocol - [TMN \(Tris-MgCl<sub>2</sub>-NaCl buffer\) \(cshlp.org\)](#)
10. Check the Protocol for fixation, acetylation, and dehydration
11. The required Proteinase K concentration depends largely on the tissue type. By default, we use 5µl/100ml of proteinase K for E14.5 mouse embryo and 35-40 µl/100ml for adult mouse, monkey or human brain.
12. Be sure to visually inspect the slides from time to time for staining under the microscope.